Amendments to the Specification

Please replace the paragraph beginning at page 6, line 19 with the following amended paragraph:

~~FIGURE 2~~ FIGURES 2A-B Binding of huPrPc from extracts of normal human brain to affinity resins in a column format. Brain homogenate and beads were prepared and equilibrated in either phosphate (PBS) or citrate phosphate dextrose (CPD) buffers. The strength of the signal on the Western blots is a function of the strength of PrPc binding to the resin. Lane 1 contains molecular weight marker (MW); Lane 2, 20 μl of 0.1% normal human brain homogenate. Lane 3-8, PrPc eluted from beads.

Please replace the paragraph beginning at page 6, line 25 with the following amended paragraph:

~~FIGURE 3~~ FIGURES 3A-B Binding of huPrPsc from extracts of CJD infected human brain to affinity resins in a batch format. The ~~figure is a Western blot that shows~~ figures are Western blots that show the amount of prion eluted from beads following contact with a homogenate containing huPrPsc from a patient with sporadic CJD. The beads were washed following contact with the homogenate that were either treated with PK to reveal the presence of PrPres or remained untreated. They were boiled in buffer containing SDS to release bound protein, and the samples were resolved by SDS-PAGE followed by Western blotting. The binding of huPrPsc and PrPc to the resins is demonstrated by presence of PrP specific bands following probing with a monoclonal antibody, 3F4. Peptide sequences are indicated at the top of the gel. Samples digested with PK are identified as (+), undigested as (-).

Please replace the paragraph beginning at page 7, line 3 with the following amended paragraph:

~~FIGURE 4~~ FIGURES 4A-B Binding of huPrPsc from extracts of CJD infected human brain to affinity resins in a column format. Peptide sequences are indicated at the top of the gel. Samples previously digested with PK are identified as + and undigested as

-. Controls included 20µl of 1% brain homogenate. PrPc and PrPsc were specifically detected using monoclonal antibody 3F4 and visualized by detection of a chemiluminescent signal.

Please replace the paragraph beginning at page 7, line 11 with the following amended paragraph:

~~FIGURE 6~~ FIGURES 6A-F Removal of PrPres from infected RBCCs by various affinity resins. Red Blood Cell Concentrates (RBCCs) were spiked with brain homogenate from hamsters infected with Scrapie and passed in succession through columns of resins with various affinity ligands. Resin-bound proteins were analyzed by gel electrophoresis. Gel loading pattern is shown in Table 11.

Please replace the paragraph beginning at page 46, line 7 with the following amended paragraph:

Binding of PrPc from normal human brain to trimer resins is shown in ~~Figure 2~~ Figures 2A-B. Ten mg of each resin (Amino, HYD (SEQ ID NO:206)), RWD (SEQ ID NO:113), SYA (SEQ ID NO:108), SYF (SEQ ID NO:213), and YEY (SEQ ID NO:154)), per column was used. The amino resin is the base polymer from which the peptides are synthesized and has some affinity to prion protein. Resins were equilibrated with either PBS, or CPD at pH 7.4. Frozen normal human brain tissue was used as the source of huPrPc. It was first thawed on wet ice. A sample of 10% brain homogenate prepared in PBS or in CPD was solubilized with 1% Sarcosyl and clarified by centrifugation at 14,000 rpm for five minutes. The supenatant was recovered and diluted 100 times to a final concentration of brain homogenate and Sarcosyl of 0.1% and 0.01%, respectively. One milliliter of this material was applied to the column and the flow through was collected. Beads were washed with 20 ml of PBS or CPD, and 1 mg of beads (dry weight) was used for evaluation of PrPc binding by Western blot as described below.

Please replace the paragraph beginning at page 47, line 12 with the following amended paragraph:

Binding of PrPsc in a brain homogenate taken from a sporadic CJD patient to resins in a flow-through format is shown in ~~Figure 4~~ Figures 4A-B. Fifty milligrams of each resin (Amino, RWERED (SEQ ID NO:157), LW (SEQ ID NO:50), EYY (SEQ ID NO:214), HYD (SEQ ID NO:206)), RWD (SEQ ID NO:113), SYA (SEQ ID NO:108), SYF (SEQ ID NO:213), and YEY (SEQ ID NO:154)) was used in experiment. The Captiva 96-well Filter Plate (CaptiVac Vacuum Sistem, ANSYS Technologies, Inc, Cat.# 796) was used instead of individual columns. Resins were prepared according to the general protocol described above. Resins were equilibrated with CPD at pH 7.4. Frozen brain tissue from a sporadic CJD patient was used as the source of huPrPc and huPrPsc. A sample of 10% brain homogenate was prepared in CPD treated with 1% sarcosyl and clarified by centrifugation at 14,000 rpm for five minutes. The supernatant was recovered and diluted ten times to give a final concentration of brain homogenate and sarcosyl of 1% and 0.1 % respectively. To each well, 250 µl of this material was applied . The material was allowed to pass through the resin under gravity with a contact time of about four minutes and flow through was collected. Resins were washed with 2.5 ml of CPD. One milligram of beads (dry weight) was incubated with PK (100 µg/ml) at 37°C for one hour. The usual processing of the beads for Western blot followed, as described above.